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(54) Title: SECRETED AND TRANSMEMBRANE POLYPEPTIDES AND NUCLEIC ACIDS ENCODING THE SAME

(57) Abstract

The present invention is directed to novel polypeptides and to nucleic acid molecules encoding those polypeptides. Also provided herein are vectors and host cells comprising those nucleic acid sequences, chimeric polypeptides molecules comprising the polypeptides of the present invention fused to heterologous polypeptide sequences, antibodies which bind to the polypeptides of the present invention and to methods for producing the polypeptides of the present invention.

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- 74(1):111-116 (July 1995), reporting that platelets have leucine rich repeats. Another protein of particular interest which has been reported to have leucine-rich repeats is the SLIT protein which has been reported to be useful in treating neuro-degenerative diseases such as Alzheimer's disease, nerve damage such as in Parkinson's disease, and for diagnosis of cancer, see, Artavanis-Tsakonas, S. and Rothberg, J. M., WO9210518-A1 by Yale University. Other studies reporting on the biological functions of proteins having leucine-rich repeats include: Tayar, N., et al., Mol. Cell Endocrinol., (Ireland), 125(1-2):65-70 (Dec. 1996) (gonadotropin receptor involvement); Miura, Y., et al., Nippon Rinsho (Japan), 54(7):1784-1789 (July 1996) (apoptosis involvement); Harris, P. C., et al., J. Am. Soc. Nephrol., 6(4):1125-1133 (Oct. 1995) (kidney disease involvement); and Ruoslahti, E. I., et al., WO9110727-A by La Jolla Cancer Research Foundation (decorin binding to transforming growth factor β involvement for treatment for cancer, wound healing and scarring).
- 10 Efforts are therefore being undertaken by both industry and academia to identify new proteins having leucine rich repeats to better understand protein-protein interactions, neuronal development and adhesin molecules. Of particular interest are those proteins having leucine rich repeats and homology to known proteins having leucine rich repeats such as the SLIT protein. We herein describe novel polypeptides having homology to SLIT, designated herein as PRO266 polypeptides.

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14. **PRO269**

Thrombomodulin binds to and regulates the activity of thrombin. It is important in the control of blood coagulation. Thrombomodulin functions as a natural anticoagulant by accelerating the activation of protein C by thrombin. Soluble thrombomodulin may have therapeutic use as an antithrombotic agent with reduced risk for hemorrhage as compared with heparin. Thrombomodulin is a cell surface trans-membrane glycoprotein, present on endothelial cells and platelets. A smaller, functionally active form of thrombomodulin circulates in the plasma and is also found in urine. (In Haeberli, A., Human Protein Data, VCH Oub., N.Y., 1992). Peptides having homology to thrombomodulin are particularly desirable.

20 We herein describe the identification and characterization of novel polypeptides having homology to thrombomodulin, designated herein as PRO269 polypeptides.

15. **PRO287**

30 Procollagen C-proteinase enhancer protein binds to and enhances the activity of bone morphogenic protein "BMP1"/procollagen C-proteinase (PCP). It plays a role in extracellular matrix deposition. BMP1 proteins may be used to induce bone and/or cartilage formation and in wound healing and tissue repair. Therefore, procollagen C-proteinase enhancer protein, BMP1 and proteins having homology thereto, are of interest to the scientific and medical communities.

35 We herein describe the identification and characterization of novel polypeptides having homology to procollagen C-proteinase enhancer protein precursor and procollagen C-proteinase enhancer protein, designated herein as PRO287 polypeptides.

12. **PRO258**

Applicants have identified a cDNA clone that encodes a novel polypeptide having homology to CRTAM and poliovirus receptor precursors, wherein the polypeptide is designated in the present application as "PRO258".

In one embodiment, the invention provides an isolated nucleic acid molecule comprising DNA encoding a PRO258 polypeptide. In one aspect, the isolated nucleic acid comprises DNA encoding the PRO258 polypeptide 5 having amino acid residues 1 to 398 of Figure 32 (SEQ ID NO:84), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions.

In another embodiment, the invention provides isolated PRO258 polypeptide. In particular, the invention provides isolated native sequence PRO258 polypeptide, which in one embodiment, includes an amino acid sequence 10 comprising residues 1 to 398 of Figure 32 (SEQ ID NO:84). An additional embodiment of the present invention is directed to an isolated extracellular domain of a PRO258 polypeptide.

13. **PRO266**

Applicants have identified a cDNA clone that encodes a novel polypeptide, wherein the polypeptide is 15 designated in the present application as "PRO266".

In one embodiment, the invention provides an isolated nucleic acid molecule comprising DNA encoding a PRO266 polypeptide. In one aspect, the isolated nucleic acid comprises DNA encoding the PRO266 polypeptide having amino acid residues 1 to 696 of Figure 34 (SEQ ID NO:91), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency 20 conditions.

In another embodiment, the invention provides isolated PRO266 polypeptide. In particular, the invention provides isolated native sequence PRO266 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 to 696 of Figure 34 (SEQ ID NO:91).

25 14. **PRO269**

Applicants have identified a cDNA clone that encodes a novel polypeptide, wherein the polypeptide is designated in the present application as PRO269.

In one embodiment, the invention provides an isolated nucleic acid molecule comprising DNA encoding a PRO269 polypeptide. In one aspect, the isolated nucleic acid comprises DNA encoding the PRO269 polypeptide 30 having amino acid residues 1 to 490 of Fig. 36 (SEQ ID NO:96), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions.

In another embodiment, the invention provides isolated PRO269 polypeptide. In particular, the invention provides isolated native sequence PRO269 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 to 490 of Figure 36 (SEQ ID NO:96). An additional embodiment of the present invention is 35 directed to an isolated extracellular domain of a PRO269 polypeptide.

Figure 27 shows a nucleotide sequence (SEQ ID NO:70) of a native sequence PRO221 cDNA, wherein SEQ ID NO:70 is a clone designated herein as "UNQ195" and/or "DNA33089-1132".

Figure 28 shows the amino acid sequence (SEQ ID NO:71) derived from the coding sequence of SEQ ID NO:70 shown in Figure 27.

5 Figure 29 shows a nucleotide sequence (SEQ ID NO:72) of a native sequence PRO227 cDNA, wherein SEQ ID NO:72 is a clone designated herein as "UNQ201" and/or "DNA33786-1132".

Figure 30 shows the amino acid sequence (SEQ ID NO:73) derived from the coding sequence of SEQ ID NO:72 shown in Figure 29.

Figure 31 shows a nucleotide sequence (SEQ ID NO:83) of a native sequence PRO258 cDNA, wherein SEQ ID NO:83 is a clone designated herein as "UNQ225" and/or "DNA35918-1174".

10 Figure 32 shows the amino acid sequence (SEQ ID NO:84) derived from the coding sequence of SEQ ID NO:83 shown in Figure 31.

Figure 33 shows a nucleotide sequence (SEQ ID NO:90) of a native sequence PRO266 cDNA, wherein SEQ ID NO:90 is a clone designated herein as "UNQ233" and/or "DNA37150-1178".

15 Figure 34 shows the amino acid sequence (SEQ ID NO:91) derived from the coding sequence of SEQ ID NO:90 shown in Figure 33.

Figure 35 shows a nucleotide sequence (SEQ ID NO:95) of a native sequence PRO269 cDNA, wherein SEQ ID NO:95 is a clone designated herein as "UNQ236" and/or "DNA38260-1180".

Figure 36 shows the amino acid sequence (SEQ ID NO:96) derived from the coding sequence of SEQ ID NO:95 shown in Figure 35.

20 Figure 37 shows a nucleotide sequence (SEQ ID NO:103) of a native sequence PRO287 cDNA, wherein SEQ ID NO:103 is a clone designated herein as "UNQ250" and/or "DNA39969-1185".

Figure 38 shows the amino acid sequence (SEQ ID NO:104) derived from the coding sequence of SEQ ID NO:103 shown in Figure 37.

25 Figure 39 shows a nucleotide sequence (SEQ ID NO:108) of a native sequence PRO214 cDNA, wherein SEQ ID NO:108 is a clone designated herein as "UNQ188" and/or "DNA32286-1191".

Figure 40 shows the amino acid sequence (SEQ ID NO:109) derived from the coding sequence of SEQ ID NO:108 shown in Figure 39.

Figure 41 shows a nucleotide sequence (SEQ ID NO:113) of a native sequence PRO317 cDNA, wherein SEQ ID NO:113 is a clone designated herein as "UNQ278" and/or "DNA33461-1199".

30 Figure 42 shows the amino acid sequence (SEQ ID NO:114) derived from the coding sequence of SEQ ID NO:113 shown in Figure 41.

Figure 43 shows a nucleotide sequence (SEQ ID NO:118) of a native sequence PRO301 cDNA, wherein SEQ ID NO:118 is a clone designated herein as "UNQ264" and/or "DNA40628-1216".

35 Figure 44 shows the amino acid sequence (SEQ ID NO:119) derived from the coding sequence of SEQ ID NO:118 shown in Figure 43.

Figure 45 shows a nucleotide sequence (SEQ ID NO:126) of a native sequence PRO224 cDNA, wherein SEQ ID NO:126 is a clone designated herein as "UNQ198" and/or "DNA33221-1133".

PRO227 has amino acid identity with the amino acid sequence of platelet glycoprotein V precursor. The same results were obtained for human glycoprotein V. Different portions of these two proteins show the following percent identities of 30%, 28%, 28%, 31%, 35%, 39% and 27%.

Accordingly, it is presently believed that PRO220, PRO221 and PRO227 polypeptides disclosed in the present application are newly identified members of the leucine rich repeat protein superfamily and that each 5 possesses protein-protein binding capabilities typical of the leucine rich repeat protein superfamily. It is also believed that they have capabilities similar to those of SLIT, the leucine rich repeat protein and human glycoprotein V.

12. Full-length PRO258 Polypeptides

The present invention provides newly identified and isolated nucleotide sequences encoding polypeptides 10 referred to in the present application as PRO258. In particular, Applicants have identified and isolated cDNA encoding a PRO258 polypeptide, as disclosed in further detail in the Examples below. Using BLAST and FastA sequence alignment computer programs, Applicants found that various portions of the PRO258 polypeptide have significant homology with the CRTAM and poliovirus receptors. Accordingly, it is presently believed that PRO258 polypeptide disclosed in the present application is a newly identified member of the Ig superfamily and possesses virus 15 receptor capabilities or regulates immune function as typical of this family.

13. Full-length PRO266 Polypeptides

The present invention provides newly identified and isolated nucleotide sequences encoding polypeptides referred to in the present application as PRO266. In particular, Applicants have identified and isolated cDNA 20 encoding a PRO266 polypeptide, as disclosed in further detail in the Examples below. Using BLAST and FastA sequence alignment computer programs, Applicants found that various portions of the PRO266 polypeptide have significant homology with the SLIT protein from Drosophila. Accordingly, it is presently believed that PRO266 polypeptide disclosed in the present application is a newly identified member of the leucine rich repeat family and possesses ligand-ligand binding activity and neuronal development typical of this family. SLIT has been shown to 25 be useful in the study and treatment of Alzheimer's disease, *supra*, and thus, PRO266 may have involvement in the study and cure of this disease.

14. Full-length PRO269 Polypeptides

The present invention provides newly identified and isolated nucleotide sequences encoding polypeptides 30 referred to in the present application as PRO269. In particular, Applicants have identified and isolated cDNA encoding a PRO269 polypeptide, as disclosed in further detail in the Examples below. Using BLAST, FastA and sequence alignment computer programs, Applicants found that the amino acid sequence encoded by nucleotides 314 to 1783 of the full-length native sequence PRO269 (shown in Figure 35 and SEQ ID NO:95) has significant homology to human urinary thrombomodulin and various thrombomodulin analogues respectively, to which it was aligned. 35 Accordingly, it is presently believed that PRO269 polypeptide disclosed in the present application is a newly identified member of the thrombomodulin family.

conservative amino acid replacements. Insertions or deletions may optionally be in the range of 1 to 5 amino acids. The variation allowed may be determined by systematically making insertions, deletions or substitutions of amino acids in the sequence and testing the resulting variants for activity in the *in vitro* assay described in the Examples below.

The variations can be made using methods known in the art such as oligonucleotide-mediated (site-directed) 5 mutagenesis, alanine scanning, and PCR mutagenesis. Site-directed mutagenesis [Carter et al., *Nucl. Acids Res.*, 13:4331 (1986); Zoller et al., *Nucl. Acids Res.*, 10:6487 (1987)], cassette mutagenesis [Wells et al., *Gene*, 34:315 (1985)], restriction selection mutagenesis [Wells et al., *Philos. Trans. R. Soc. London SerA*, 317:415 (1986)] or other known techniques can be performed on the cloned DNA to produce the desired PRO polypeptide variant DNA.

Scanning amino acid analysis can also be employed to identify one or more amino acids along a contiguous 10 sequence. Among the preferred scanning amino acids are relatively small, neutral amino acids. Such amino acids include alanine, glycine, serine, and cysteine. Alanine is typically a preferred scanning amino acid among this group because it eliminates the side-chain beyond the beta-carbon and is less likely to alter the main-chain conformation of the variant. Alanine is also typically preferred because it is the most common amino acid. Further, it is frequently found in both buried and exposed positions [Creighton, *The Proteins*, (W.H. Freeman & Co., N.Y.); Chothia, *L. 15 Mol. Biol.*, 150:1 (1976)]. If alanine substitution does not yield adequate amounts of variant, an isoteric amino acid can be used.

51. Modifications of PRO Polypeptides

Covalent modifications of PRO polypeptides are included within the scope of this invention. One type of 20 covalent modification includes reacting targeted amino acid residues of the PRO polypeptide with an organic derivatizing agent that is capable of reacting with selected side chains or the N- or C-terminal residues of the PRO polypeptide. Derivatization with bifunctional agents is useful, for instance, for crosslinking a PRO polypeptide to a water-insoluble support matrix or surface for use in the method for purifying anti-PRO polypeptide antibodies, and vice-versa. Commonly used crosslinking agents include, e.g., 1,1-bis(diazoacetyl)-2-phenylethane, glutaraldehyde, 25 N-hydroxysuccinimide esters, for example, esters with 4-azidosalicylic acid, homobifunctional imidoesters, including disuccinimidyl esters such as 3,3'-dithiobis(succinimidylpropionate), bifunctional maleimides such as bis-N-maleimido-1,8-octane and agents such as methyl-3-[(*p*-azidophenyl)dithio]propioimidate.

Other modifications include deamidation of glutaminyl and asparaginyl residues to the corresponding 30 glutamyl and aspartyl residues, respectively, hydroxylation of proline and lysine, phosphorylation of hydroxyl groups of seryl or threonyl residues, methylation of the α -amino groups of lysine, arginine, and histidine side chains [T.E. Creighton, *Proteins: Structure and Molecular Properties*, W.H. Freeman & Co., San Francisco, pp. 79-86 (1983)], acetylation of the N-terminal amine, and amidation of any C-terminal carboxyl group.

Another type of covalent modification of the PRO polypeptides included within the scope of this invention comprises altering the native glycosylation pattern of the polypeptide. "Altering the native glycosylation pattern" is 35 intended for purposes herein to mean deleting one or more carbohydrate moieties found in a native sequence PRO polypeptide, and/or adding one or more glycosylation sites that are not present in the native sequence PRO polypeptide.

Addition of glycosylation sites to the PRO polypeptide may be accomplished by altering the amino acid sequence. The alteration may be made, for example, by the addition of, or substitution by, one or more serine or threonine residues to the native sequence PRO polypeptide (for O-linked glycosylation sites). The PRO polypeptide amino acid sequence may optionally be altered through changes at the DNA level, particularly by mutating the DNA encoding the PRO polypeptide at preselected bases such that codons are generated that will translate into the desired 5 amino acids.

Another means of increasing the number of carbohydrate moieties on the PRO polypeptide polypeptide is by chemical or enzymatic coupling of glycosides to the polypeptide. Such methods are described in the art, e.g., in WO 87/05330 published 11 September 1987, and in Aplin and Wriston, CRC Crit. Rev. Biochem., pp. 259-306 (1981).

10 Removal of carbohydrate moieties present on the PRO polypeptide may be accomplished chemically or enzymatically or by mutational substitution of codons encoding for amino acid residues that serve as targets for glycosylation. Chemical deglycosylation techniques are known in the art and described, for instance, by Hakimuddin, et al., Arch. Biochem. Biophys., 259:52 (1987) and by Edge et al., Anal. Biochem., 118:131 (1981). Enzymatic cleavage of carbohydrate moieties on polypeptides can be achieved by the use of a variety of endo- and exo-glycosidases as described by Thotakura et al., Meth. Enzymol., 138:350 (1987).

Another type of covalent modification of PRO polypeptides of the invention comprises linking the PRO polypeptide to one of a variety of nonproteinaceous polymers, e.g., polyethylene glycol, polypropylene glycol, or polyoxyalkylenes, in the manner set forth in U.S. Patent Nos. 4,640,835; 4,496,689; 4,301,144; 4,670,417; 4,791,192 or 4,179,337.

20 The PRO polypeptides of the present invention may also be modified in a way to form a chimeric molecule comprising a PRO polypeptide fused to another, heterologous polypeptide or amino acid sequence. In one embodiment, such a chimeric molecule comprises a fusion of the PRO polypeptide with a tag polypeptide which provides an epitope to which an anti-tag antibody can selectively bind. The epitope tag is generally placed at the amino- or carboxyl- terminus of the PRO polypeptide. The presence of such epitope-tagged forms of the PRO 25 polypeptide can be detected using an antibody against the tag polypeptide. Also, provision of the epitope tag enables the PRO polypeptide to be readily purified by affinity purification using an anti-tag antibody or another type of affinity matrix that binds to the epitope tag. In an alternative embodiment, the chimeric molecule may comprise a fusion of the PRO polypeptide with an immunoglobulin or a particular region of an immunoglobulin. For a bivalent form of the chimeric molecule, such a fusion could be to the Fc region of an IgG molecule.

30 Various tag polypeptides and their respective antibodies are well known in the art. Examples include poly-histidine (poly-his) or poly-histidine-glycine (poly-his-gly) tags; the flu HA tag polypeptide and its antibody 12CAS [Field et al., Mol. Cell. Biol., 8:2159-2165 (1988)]; the c-myc tag and the 8F9, 3C7, 6E10, G4, B7 and 9E10 antibodies thereto [Evan et al., Molecular and Cellular Biology, 5:3610-3616 (1985)]; and the Herpes Simplex virus glycoprotein D (gD) tag and its antibody [Paborsky et al., Protein Engineering, 3(6):547-553 (1990)]. Other tag 35 polypeptides include the Flag-peptide [Hopp et al., BioTechnology, 6:1204-1210 (1988)]; the KT3 epitope peptide [Martin et al., Science, 255:192-194 (1992)]; an α -tubulin epitope peptide [Skinner et al., J. Biol. Chem., 266:15163-15166 (1991)]; and the T7 gene 10 protein peptide tag [Lutz-Freyermuth et al., Proc. Natl. Acad. Sci. USA, 87:6393-

6397 (1990)].

52. Modification of PRO317

Amino acid sequence variants of PRO317 are prepared by introducing appropriate nucleotide changes into the PRO317 DNA, or by *in vitro* synthesis of the desired PRO317 polypeptide. Such variants include, for example, 5 deletions from, or insertions or substitutions of, residues within the amino acid sequence shown for human PRO317 in Figure 42. Any combination of deletion, insertion, and substitution is made to arrive at the final construct, provided that the final construct possesses the desired characteristics. The amino acid changes also may alter post-translational processes of the PRO317, such as changing the number or position of glycosylation sites. Moreover, like most mammalian genes, PRO317 is presumably encoded by multi-exon genes. Alternative mRNA constructs 10 which may be attributed to different mRNA splicing events following transcription, and which share large regions of identity with the cDNAs claimed herein, are considered to be within the scope of the present invention.

For the design of amino acid sequence variants of PRO317, the location of the mutation site and the nature of the mutation will depend on the PRO317 characteristic(s) to be modified. For example, candidate PRO317 antagonists or agonists will be initially selected by locating sites that are identical or highly conserved among 15 PRO317, EBAF-1, LEFTY, and other members of the TGF- superfamily. The sites for mutation can be modified individually or in series, *e.g.*, by (1) substituting first with conservative amino acid choices and then with more radical selections depending upon the results achieved, (2) deleting the target residue, or (3) inserting residues of the same or a different class adjacent to the located site, or combinations of options 1-3.

A useful method for identification of certain residues or regions of the PRO317 polypeptide that are 20 preferred locations for mutagenesis is called "alanine scanning mutagenesis," as described by Cunningham and Wells, Science, 244: 1081-1085 (1989). Here, a residue or group of target residues are identified (*e.g.*, charged residues such as arg, asp, his, lys, and glu) and replaced by a neutral or negatively charged amino acid (most preferably alanine or polyalanine) to affect the interaction of the amino acids with the surrounding aqueous environment in or outside the cell. Those domains demonstrating functional sensitivity to the substitutions then are refined by 25 introducing further or other variants at or for the sites of substitution. Thus, while the site for introducing an amino acid sequence variation is predetermined, the nature of the mutation *per se* need not be predetermined. For example, to optimize the performance of a mutation at a given site, alanine scanning or random mutagenesis is conducted at the target codon or region and the PRO317 variants produced are screened for the optimal combination of desired activity.

30 There are two principal variables in the construction of amino acid sequence variants: the location of the mutation site and the nature of the mutation. These are variants from the Figure 42 sequence, and may represent naturally occurring alleles (which will not require manipulation of the PRO317 DNA) or predetermined mutant forms made by mutating the DNA, either to arrive at an allele or a variant not found in nature. In general, the location and nature of the mutation chosen will depend upon the PRO317 characteristic to be modified.

35 Amino acid sequence deletions generally range from about 1 to 30 residues, more preferably about 1 to 10 residues, and typically are contiguous. Contiguous deletions ordinarily are made in even numbers of residues, but single or odd numbers of deletions are within the scope hereof. Deletions may be introduced into regions of low

(Field *et al.*, Mol. Cell. Biol., **8**:2159-2165 (1988)); the c-myc tag and the 8F9, 3C7, 6E10, G4, B7, and 9E10 antibodies thereto (Evan *et al.*, Molecular and Cellular Biology, **5**:3610-3616 (1985)); and the Herpes Simplex virus glycoprotein D (gD) tag and its antibody (Paborsky *et al.*, Protein Engineering, **3**(6):547-553 (1990)). Other tag polypeptides include the Flag-peptide (Hopp *et al.*, Bio/Technology, **6**:1204-1210 (1988)); the KT3 epitope peptide (Martin *et al.*, Science, **255**:192-194 (1992)); an "-tubulin epitope peptide (Skinner *et al.*, J. Biol. Chem., **266**:15163-5 15166 (1991)); and the T7 gene 10 protein peptide tag (Lutz-Freyermuth *et al.*, Proc. Natl. Acad. Sci. USA, **87**:6393-6397 (1990)).

53. Preparation of PRO Polypeptides

The description below relates primarily to production of PRO polypeptides by culturing cells transformed 10 or transfected with a vector containing the desired PRO polypeptide nucleic acid. It is, of course, contemplated that alternative methods, which are well known in the art, may be employed to prepare the PRO polypeptide. For instance, the PRO polypeptide sequence, or portions thereof, may be produced by direct peptide synthesis using solid-phase techniques [see, e.g., Stewart *et al.*, Solid-Phase Peptide Synthesis, W.H. Freeman Co., San Francisco, CA 15 (1969); Merrifield, J. Am. Chem. Soc., **85**:2149-2154 (1963)]. *In vitro* protein synthesis may be performed using manual techniques or by automation. Automated synthesis may be accomplished, for instance, using an Applied Biosystems Peptide Synthesizer (Foster City, CA) using manufacturer's instructions. Various portions of the desired PRO polypeptide may be chemically synthesized separately and combined using chemical or enzymatic methods to produce the full-length PRO polypeptide.

20 A. Isolation of DNA Encoding PRO Polypeptides

DNA encoding PRO polypeptides may be obtained from a cDNA library prepared from tissue believed to possess the desired PRO polypeptide mRNA and to express it at a detectable level. Accordingly, human PRO polypeptide DNA can be conveniently obtained from a cDNA library prepared from human tissue, such as described in the Examples. The PRO polypeptide-encoding gene may also be obtained from a genomic library or by 25 oligonucleotide synthesis.

Libraries can be screened with probes (such as antibodies to the desired PRO polypeptide or oligonucleotides of at least about 20-80 bases) designed to identify the gene of interest or the protein encoded by it. Screening the cDNA or genomic library with the selected probe may be conducted using standard procedures, such as described in Sambrook *et al.*, Molecular Cloning: A Laboratory Manual (New York: Cold Spring Harbor Laboratory Press, 30 1989). An alternative means to isolate the gene encoding the desired PRO polypeptide is to use PCR methodology [Sambrook *et al.*, supra; Dieffenbach *et al.*, PCR Primer: A Laboratory Manual (Cold Spring Harbor Laboratory Press, 1995)].

The Examples below describe techniques for screening a cDNA library. The oligonucleotide sequences selected as probes should be of sufficient length and sufficiently unambiguous that false positives are minimized. The 35 oligonucleotide is preferably labeled such that it can be detected upon hybridization to DNA in the library being screened. Methods of labeling are well known in the art, and include the use of radiolabels like ³²P-labeled ATP, biotinylation or enzyme labeling. Hybridization conditions, including moderate stringency and high stringency, are

provided in Sambrook et al., *supra*.

Sequences identified in such library screening methods can be compared and aligned to other known sequences deposited and available in public databases such as GenBank or other private sequence databases. Sequence identity (at either the amino acid or nucleotide level) within defined regions of the molecule or across the full-length sequence can be determined through sequence alignment using computer software programs such as
5 BLAST, ALIGN, DNAsstar, and INHERIT which employ various algorithms to measure homology.

Nucleic acid having protein coding sequence may be obtained by screening selected cDNA or genomic libraries using the deduced amino acid sequence disclosed herein for the first time, and, if necessary, using conventional primer extension procedures as described in Sambrook et al., *supra*, to detect precursors and processing intermediates of mRNA that may not have been reverse-transcribed into cDNA.

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B. Selection and Transformation of Host Cells

Host cells are transfected or transformed with expression or cloning vectors described herein for PRO polypeptide production and cultured in conventional nutrient media modified as appropriate for inducing promoters, selecting transformants, or amplifying the genes encoding the desired sequences. The culture conditions, such as
15 media, temperature, pH and the like, can be selected by the skilled artisan without undue experimentation. In general, principles, protocols, and practical techniques for maximizing the productivity of cell cultures can be found in Mammalian Cell Biotechnology: a Practical Approach, M. Butler, ed. (IRL Press, 1991) and Sambrook et al., *supra*.

Methods of transfection are known to the ordinarily skilled artisan, for example, CaPO₄ and electroporation.

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Depending on the host cell used, transformation is performed using standard techniques appropriate to such cells. The calcium treatment employing calcium chloride, as described in Sambrook et al., *supra*, or electroporation is generally used for prokaryotes or other cells that contain substantial cell-wall barriers. Infection with *Agrobacterium tumefaciens* is used for transformation of certain plant cells, as described by Shaw et al., *Gene*, 23:315 (1983) and WO 89/05859 published 29 June 1989. For mammalian cells without such cell walls, the calcium phosphate precipitation method of Graham and van der Eb, *Virology*, 52:456-457 (1978) can be employed. General aspects of mammalian cell host system transformations have been described in U.S. Patent No. 4,399,216. Transformations into yeast are typically carried out according to the method of Van Solingen et al., *J. Bact.*, 130:946 (1977) and Hsiao et al., *Proc. Natl. Acad. Sci. (USA)*, 76:3829 (1979). However, other methods for introducing DNA into cells, such as by nuclear microinjection, electroporation, bacterial protoplast fusion with intact cells, or polycations, e.g.,
25 polybrene, polyornithine, may also be used. For various techniques for transforming mammalian cells, see Keown et al., Methods in Enzymology, 185:527-537 (1990) and Mansour et al., *Nature*, 336:348-352 (1988).

30

Suitable host cells for cloning or expressing the DNA in the vectors herein include prokaryote, yeast, or higher eukaryote cells. Suitable prokaryotes include but are not limited to eubacteria, such as Gram-negative or Gram-positive organisms, for example, Enterobacteriaceae such as *E. coli*. Various *E. coli* strains are publicly
35 available, such as *E. coli* K12 strain MM294 (ATCC 31,446); *E. coli* X1776 (ATCC 31,537); *E. coli* strain W3110 (ATCC 27,325) and K5 772 (ATCC 53,635). Other suitable prokaryotic host cells include Enterobacteriaceae such as *Escherichia*, e.g., *E. coli*, *Enterobacter*, *Erwinia*, *Klebsiella*, *Proteus*, *Salmonella*, e.g., *Salmonella typhimurium*,

Serratia, e.g., *Serratia marcescens*, and *Shigella*, as well as *Bacilli* such as *B. subtilis* and *B. licheniformis* (e.g., *B. licheniformis* 41P disclosed in DD 266,710 published 12 April 1989), *Pseudomonas* such as *P. aeruginosa*, and *Streptomyces*. Various *E. coli* strains are publicly available, such as *E. coli* K12 strain MM294 (ATCC 31,446); *E. coli* X1776 (ATCC 31,537); *E. coli* strain W3110 (ATCC 27,325); and K5 772 (ATCC 53,635). These examples are illustrative rather than limiting. Strain W3110 is one particularly preferred host or parent host because it is a common host strain for recombinant DNA product fermentations. Preferably, the host cell secretes minimal amounts of proteolytic enzymes. For example, strain W3110 may be modified to effect a genetic mutation in the genes encoding proteins endogenous to the host, with examples of such hosts including *E. coli* W3110 strain 1A2, which has the complete genotype *tonA*; *E. coli* W3110 strain 9E4, which has the complete genotype *tonA ptr3*; *E. coli* W3110 strain 27C7 (ATCC 55,244), which has the complete genotype *tonA ptr3 phoA E15 (argF-lac)169 degP*; *ompT kan'*; *E. coli* W3110 strain 37D6, which has the complete genotype *tonA ptr3 phoA E15 (argF-lac)169 degP ompT rbs7 ilvG kan'*; *E. coli* W3110 strain 40B4, which is strain 37D6 with a non-kanamycin resistant *degP* deletion mutation; and an *E. coli* strain having mutant periplasmic protease disclosed in U.S. Patent No. 4,946,783 issued 7 August 1990. Alternatively, *in vitro* methods of cloning, e.g., PCR or other nucleic acid polymerase reactions, are suitable.

In addition to prokaryotes, eukaryotic microbes such as filamentous fungi or yeast are suitable cloning or expression hosts for PRO polypeptide-encoding vectors. *Saccharomyces cerevisiae* is a commonly used lower eukaryotic host microorganism. Others include *Schizosaccharomyces pombe* (Beach and Nurse, *Nature*, **290**: 140 [1981]; EP 139,383 published 2 May 1985); *Kluyveromyces* hosts (U.S. Patent No. 4,943,529; Fleer et al., *Bio/Technology*, **9**: 968-975 (1991)) such as, e.g., *K. lactis* (MW98-8C, CBS683, CBS4574; Louvencourt et al., *J. Bacteriol.*, **73** [1983]), *K. fragilis* (ATCC 12,424), *K. bulgaricus* (ATCC 16,045), *K. wickeramii* (ATCC 24,178), *K. waltii* (ATCC 56,500), *K. drosophilae* (ATCC 36,906; Van den Berg et al., *Bio/Technology*, **8**: 135 (1990)), *K. thermotolerans*, and *K. marxianus*; *yarrowiae* (EP 402,226); *Pichia pastoris* (EP 183,070; Sreekrishna et al., *J. Basic Microbiol.*, **28**: 265-278 [1988]); *Candida*; *Trichoderma reesiae* (EP 244,234); *Neurospora crassa* (Case et al., *Proc. Natl. Acad. Sci. USA*, **76**: 5259-5263 [1979]); *Schwanniomyces* such as *Schwanniomyces occidentalis* (EP 394,538 published 31 October 1990); and filamentous fungi such as, e.g., *Neurospora*, *Penicillium*, *Tolypocladium* (WO 91/00357 published 10 January 1991), and *Aspergillus* hosts such as *A. nidulans* (Ballance et al., *Biochem. Biophys. Res. Commun.*, **112**: 284-289 [1983]; Tilburn et al., *Gene*, **26**: 205-221 [1983]; Yelton et al., *Proc. Natl. Acad. Sci. USA*, **81**: 1470-1474 [1984]) and *A. niger* (Kelly and Hynes, *EMBO J.*, **4**: 475-479 [1985]). Methylotropic yeasts are suitable herein and include, but are not limited to, yeast capable of growth on methanol selected from the genera consisting of *Hansenula*, *Candida*, *Kloeckera*, *Pichia*, *Saccharomyces*, *Torulopsis*, and *Rhodotorula*. A list of specific species that are exemplary of this class of yeasts may be found in C. Anthony, *The Biochemistry of Methylotrophs*, 269 (1982).

Suitable host cells for the expression of glycosylated PRO polypeptides are derived from multicellular organisms. Examples of invertebrate cells include insect cells such as *Drosophila S2* and *Spodoptera Sf9*, as well as plant cells. Examples of useful mammalian host cell lines include Chinese hamster ovary (CHO) and CO₂ cells. More specific examples include monkey kidney CV1 line transformed by SV40 (COS-7, ATCC CRL 1651); human embryonic kidney line (293 or 293 cells subcloned for growth in suspension culture, Graham et al., *J. Gen Virol.*,

36:59 (1977)); Chinese hamster ovary cells/-DHFR (CHO, Urlaub and Chasin, *Proc. Natl. Acad. Sci. USA*, **77**:4216 (1980)); mouse sertoli cells (TM4, Mather, *Biol. Reprod.*, **23**:243-251 (1980)); human lung cells (W138, ATCC CCL 75); human liver cells (Hep G2, HB 8065); and mouse mammary tumor (MMT 060562, ATCC CCL51). The selection of the appropriate host cell is deemed to be within the skill in the art.

5 C. Selection and Use of a Replicable Vector

The nucleic acid (*e.g.*, cDNA or genomic DNA) encoding a desired PRO polypeptide may be inserted into a replicable vector for cloning (amplification of the DNA) or for expression. Various vectors are publicly available. The vector may, for example, be in the form of a plasmid, cosmid, viral particle, or phage. The appropriate nucleic acid sequence may be inserted into the vector by a variety of procedures. In general, DNA is inserted into an 10 appropriate restriction endonuclease site(s) using techniques known in the art. Vector components generally include, but are not limited to, one or more of a signal sequence, an origin of replication, one or more marker genes, an enhancer element, a promoter, and a transcription termination sequence. Construction of suitable vectors containing one or more of these components employs standard ligation techniques which are known to the skilled artisan.

15 The PRO polypeptide of interest may be produced recombinantly not only directly, but also as a fusion polypeptide with a heterologous polypeptide, which may be a signal sequence or other polypeptide having a specific cleavage site at the N-terminus of the mature protein or polypeptide. In general, the signal sequence may be a component of the vector, or it may be a part of the PRO polypeptide DNA that is inserted into the vector. The signal sequence may be a prokaryotic signal sequence selected, for example, from the group of the alkaline phosphatase, penicillinase, lpp, or heat-stable enterotoxin II leaders. For yeast secretion the signal sequence may be, *e.g.*, the 20 yeast invertase leader, alpha factor leader (including *Saccharomyces* and *Kluyveromyces* α -factor leaders, the latter described in U.S. Patent No. 5,010,182), or acid phosphatase leader, the *C. albicans* glucoamylase leader (EP 362,179 published 4 April 1990), or the signal described in WO 90/13646 published 15 November 1990. In mammalian cell expression, mammalian signal sequences may be used to direct secretion of the protein, such as signal sequences from secreted polypeptides of the same or related species, as well as viral secretory leaders.

25 Both expression and cloning vectors contain a nucleic acid sequence that enables the vector to replicate in one or more selected host cells. Such sequences are well known for a variety of bacteria, yeast, and viruses. The origin of replication from the plasmid pBR322 is suitable for most Gram-negative bacteria, the 2μ plasmid origin is suitable for yeast, and various viral origins (SV40, polyoma, adenovirus, VSV or BPV) are useful for cloning vectors in mammalian cells.

30 Expression and cloning vectors will typically contain a selection gene, also termed a selectable marker. Typical selection genes encode proteins that (a) confer resistance to antibiotics or other toxins, *e.g.*, ampicillin, neomycin, methotrexate, or tetracycline, (b) complement auxotrophic deficiencies, or (c) supply critical nutrients not available from complex media, *e.g.*, the gene encoding D-alanine racemase for *Bacilli*.

35 An example of suitable selectable markers for mammalian cells are those that enable the identification of cells competent to take up the PRO polypeptide nucleic acid, such as DHFR or thymidine kinase. An appropriate host cell when wild-type DHFR is employed is the CHO cell line deficient in DHFR activity, prepared and propagated as described by Urlaub et al., *Proc. Natl. Acad. Sci. USA*, **77**:4216 (1980). A suitable selection gene

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for use in yeast is the *trp1* gene present in the yeast plasmid YRp7 [Stinchcomb et al., *Nature*, **282**:39 (1979); Kingsman et al., *Gene*, **1**:141 (1979); Tschemper et al., *Gene*, **10**:157 (1980)]. The *trp1* gene provides a selection marker for a mutant strain of yeast lacking the ability to grow in tryptophan, for example, ATCC No. 44076 or PEP4-1 [Jones, *Genetics*, **85**:12 (1977)].

Expression and cloning vectors usually contain a promoter operably linked to the PRO polypeptide nucleic acid sequence to direct mRNA synthesis. Promoters recognized by a variety of potential host cells are well known. 5 Promoters suitable for use with prokaryotic hosts include the β -lactamase and lactose promoter systems [Chang et al., *Nature*, **275**:615 (1978); Goeddel et al., *Nature*, **281**:544 (1979)], alkaline phosphatase, a tryptophan (*trp*) promoter system [Goeddel, *Nucleic Acids Res.*, **8**:4057 (1980); EP 36,776], and hybrid promoters such as the tac promoter [deBoer et al., *Proc. Natl. Acad. Sci. USA*, **80**:21-25 (1983)]. Promoters for use in bacterial systems also 10 will contain a Shine-Dalgarno (S.D.) sequence operably linked to the DNA encoding the desired PRO polypeptide.

Examples of suitable promoting sequences for use with yeast hosts include the promoters for 3-phosphoglycerate kinase [Hitzeman et al., *J. Biol. Chem.*, **255**:2073 (1980)] or other glycolytic enzymes [Hess et al., *J. Adv. Enzyme Reg.*, **7**:149 (1968); Holland, *Biochemistry*, **17**:4900 (1978)], such as enolase, glyceraldehyde-3-phosphate dehydrogenase, hexokinase, pyruvate decarboxylase, phosphofructokinase, glucose-6-phosphate isomerase, 15 3-phosphoglycerate mutase, pyruvate kinase, triosephosphate isomerase, phosphoglucose isomerase, and glucokinase.

Other yeast promoters, which are inducible promoters having the additional advantage of transcription controlled by growth conditions, are the promoter regions for alcohol dehydrogenase 2, isocytchrome C, acid phosphatase, degradative enzymes associated with nitrogen metabolism, metallothionein, glyceraldehyde-3-phosphate dehydrogenase, and enzymes responsible for maltose and galactose utilization. Suitable vectors and promoters for 20 yeast expression are further described in EP 73,657.

PRO polypeptide transcription from vectors in mammalian host cells is controlled, for example, by promoters obtained from the genomes of viruses such as polyoma virus, fowlpox virus (UK 2,211,504 published 5 July 1989), adenovirus (such as Adenovirus 2), bovine papilloma virus, avian sarcoma virus, cytomegalovirus, a retrovirus, hepatitis-B virus and Simian Virus 40 (SV40), from heterologous mammalian promoters, e.g., the actin promoter or an immunoglobulin promoter, and from heat-shock promoters, provided such promoters are compatible 25 with the host cell systems.

Transcription of a DNA encoding the desired PRO polypeptide by higher eukaryotes may be increased by inserting an enhancer sequence into the vector. Enhancers are cis-acting elements of DNA, usually about from 10 to 300 bp, that act on a promoter to increase its transcription. Many enhancer sequences are now known from 30 mammalian genes (globin, elastase, albumin, α -fetoprotein, and insulin). Typically, however, one will use an enhancer from a eukaryotic cell virus. Examples include the SV40 enhancer on the late side of the replication origin (bp 100-270), the cytomegalovirus early promoter enhancer, the polyoma enhancer on the late side of the replication origin, and adenovirus enhancers. The enhancer may be spliced into the vector at a position 5' or 3' to the PRO polypeptide coding sequence, but is preferably located at a site 5' from the promoter.

35 Expression vectors used in eukaryotic host cells (yeast, fungi, insect, plant, animal, human, or nucleated cells from other multicellular organisms) will also contain sequences necessary for the termination of transcription and for stabilizing the mRNA. Such sequences are commonly available from the 5' and, occasionally 3', untranslated

regions of eukaryotic or viral DNAs or cDNAs. These regions contain nucleotide segments transcribed as polyadenylated fragments in the untranslated portion of the mRNA encoding PRO polypeptides.

Still other methods, vectors, and host cells suitable for adaptation to the synthesis of PRO polypeptides in recombinant vertebrate cell culture are described in Gething et al., *Nature*, 293:620-625 (1981); Mantei et al., *Nature*, 281:40-46 (1979); EP 117,060; and EP 117,058.

5

D. Detecting Gene Amplification/Expression

Gene amplification and/or expression may be measured in a sample directly, for example, by conventional Southern blotting, Northern blotting to quantitate the transcription of mRNA [Thomas, *Proc. Natl. Acad. Sci. USA*, 77:5201-5205 (1980)], dot blotting (DNA analysis), or *in situ* hybridization, using an appropriately labeled probe, 10 based on the sequences provided herein. Alternatively, antibodies may be employed that can recognize specific duplexes, including DNA duplexes, RNA duplexes, and DNA-RNA hybrid duplexes or DNA-protein duplexes. The antibodies in turn may be labeled and the assay may be carried out where the duplex is bound to a surface, so that upon the formation of duplex on the surface, the presence of antibody bound to the duplex can be detected.

Gene expression, alternatively, may be measured by immunological methods, such as immunohistochemical 15 staining of cells or tissue sections and assay of cell culture or body fluids, to quantitate directly the expression of gene product. Antibodies useful for immunohistochemical staining and/or assay of sample fluids may be either monoclonal or polyclonal, and may be prepared in any mammal. Conveniently, the antibodies may be prepared against a native sequence PRO polypeptide or against a synthetic peptide based on the DNA sequences provided herein or against exogenous sequence fused to a PRO polypeptide DNA and encoding a specific antibody epitope.

20

E. Purification of Polypeptide

Forms of PRO polypeptides may be recovered from culture medium or from host cell lysates. If membrane-bound, it can be released from the membrane using a suitable detergent solution (e.g. Triton-X 100) or by enzymatic cleavage. Cells employed in expression of PRO polypeptides can be disrupted by various physical or chemical 25 means, such as freeze-thaw cycling, sonication, mechanical disruption, or cell lysing agents.

It may be desired to purify PRO polypeptides from recombinant cell proteins or polypeptides. The following procedures are exemplary of suitable purification procedures: by fractionation on an ion-exchange column; ethanol precipitation; reverse phase HPLC; chromatography on silica or on a cation-exchange resin such as DEAE; chromatofocusing; SDS-PAGE; ammonium sulfate precipitation; gel filtration using, for example, Sephadex G-75; 30 protein A Sepharose columns to remove contaminants such as IgG; and metal chelating columns to bind epitope-tagged forms of the PRO polypeptide. Various methods of protein purification may be employed and such methods are known in the art and described for example in Deutscher, *Methods in Enzymology*, 182 (1990); Scopes, *Protein Purification: Principles and Practice*, Springer-Verlag, New York (1982). The purification step(s) selected will depend, for example, on the nature of the production process used and the particular PRO polypeptide produced.

35

54. Uses for PRO Polypeptides

Nucleotide sequences (or their complement) encoding the PRO polypeptides of the present invention have various applications in the art of molecular biology, including uses as hybridization probes, in chromosome and gene mapping and in the generation of anti-sense RNA and DNA. PRO polypeptide-encoding nucleic acid will also be useful for the preparation of PRO polypeptides by the recombinant techniques described herein.

5 The full-length native sequence PRO polypeptide-encoding nucleic acid or portions thereof, may be used as hybridization probes for a cDNA library to isolate the full-length PRO polypeptide gene or to isolate still other genes (for instance, those encoding naturally-occurring variants of the PRO polypeptide or PRO polypeptides from other species) which have a desired sequence identity to the PRO polypeptide nucleic acid sequences. Optionally, the length of the probes will be about 20 to about 50 bases. The hybridization probes may be derived from the
10 nucleotide sequence of any of the DNA molecules disclosed herein or from genomic sequences including promoters, enhancer elements and introns of native sequence PRO polypeptide encoding DNA. By way of example, a screening method will comprise isolating the coding region of the PRO polypeptide gene using the known DNA sequence to synthesize a selected probe of about 40 bases. Hybridization probes may be labeled by a variety of labels, including radionucleotides such as ^{32}P or ^{35}S , or enzymatic labels such as alkaline phosphatase coupled to the probe via
15 avidin/biotin coupling systems. Labeled probes having a sequence complementary to that of the specific PRO polypeptide gene of the present invention can be used to screen libraries of human cDNA, genomic DNA or mRNA to determine which members of such libraries the probe hybridizes to. Hybridization techniques are described in further detail in the Examples below.

The ESTs disclosed in the present application may similarly be employed as probes, using the methods
20 disclosed herein.

The probes may also be employed in PCR techniques to generate a pool of sequences for identification of closely related PRO polypeptide sequences.

25 Nucleotide sequences encoding a PRO polypeptide can also be used to construct hybridization probes for mapping the gene which encodes that PRO polypeptide and for the genetic analysis of individuals with genetic disorders. The nucleotide sequences provided herein may be mapped to a chromosome and specific regions of a chromosome using known techniques, such as *in situ* hybridization, linkage analysis against known chromosomal markers, and hybridization screening with libraries.

30 The PRO polypeptide can be used in assays to identify its ligands. Similarly, inhibitors of the receptor/ligand binding interaction can be identified. Proteins involved in such binding interactions can also be used to screen for peptide or small molecule inhibitors or agonists of the binding interaction. Screening assays can be designed to find lead compounds that mimic the biological activity of a native PRO polypeptide or a ligand for the PRO polypeptide. Such screening assays will include assays amenable to high-throughput screening of chemical libraries, making them particularly suitable for identifying small molecule drug candidates. Small molecules contemplated include synthetic organic or inorganic compounds. The assays can be performed in a variety of formats, including protein-protein binding assays, biochemical screening assays, immunoassays and cell based assays, which are well characterized in the art.

level, DNA47412, the EST from which PCR oligos were generated to isolate the full length DNA49435-1219, has been observed to map to 11p15. Sequence homology to the 11p15 locus would indicate that PRO533 may have utility in the treatment of Usher Syndrome or Atrophia areata.

As mentioned previously, fibroblast growth factors can act upon cells in both a mitogenic and non-mitogenic manner. These factors are mitogenic for a wide variety of normal diploid mesoderm-derived and neural crest-derived 5 cells, inducing granulosa cells, adrenal cortical cells, chondrocytes, myoblasts, corneal and vascular endothelial cells (bovine or human), vascular smooth muscle cells, lens, retina and prostatic epithelial cells, oligodendrocytes, astrocytes, chondrocytes, myoblasts and osteoblasts.

Non-mitogenic actions of fibroblast growth factors include promotion of cell migration into a wound area (chemotaxis), initiation of new blood vessel formulation (angiogenesis), modulation of nerve regeneration and survival 10 (neurotrophism), modulation of endocrine functions, and stimulation or suppression of specific cellular protein expression, extracellular matrix production and cell survival. Baird, A. & Bohlen, P., *Handbook of Exp. Pharmacol.* 95(1): 369-418 (1990). These properties provide a basis for using fibroblast growth factors in therapeutic approaches to accelerate wound healing, nerve repair, collateral blood vessel formation, and the like. For example, fibroblast 15 growth factors, have been suggested to minimize myocardium damage in heart disease and surgery (U.S.P. 4,378,437).

Since the PRO245 polypeptide and nucleic acid encoding it possess sequence homology to a transmembrane protein tyrosine kinase protein and its encoding nucleic acid, probes based upon the PRO245 nucleotide sequence may be employed to identify other novel transmembrane tyrosine kinase proteins. Soluble forms of the PRO245 polypeptide may be employed as antagonists of membrane bound PRO245 activity both *in vitro* and *in vivo*. PRO245 20 polypeptides may be employed in screening assays designed to identify agonists or antagonists of the native PRO245 polypeptide, wherein such assays may take the form of any conventional cell-type or biochemical binding assay. Moreover, the PRO245 polypeptide may serve as a molecular marker for the tissues in which the polypeptide is specifically expressed.

PRO220, PRO221 and PRO227 all have leucine rich repeats. Additionally, PRO220 and PRO221 have 25 homology to SLIT and leucine rich repeat protein. Therefore, these proteins are useful in assays described in the literature, *supra*, wherein the SLIT and leucine rich repeat protein are used. Regarding the SLIT protein, PRO227 can be used in an assay to determine the affect of PRO227 on neurodegenerative disease. Additionally, PRO227 has homology to human glycoprotein V. In the case of PRO227, this polypeptide is used in an assay to determine its affect on bleeding, clotting, tissue repair and scarring.

30 The PRO266 polypeptide can be used in assays to determine if it has a role in neurodegenerative diseases or their reversal.

PRO269 polypeptides and portions thereof which effect the activity of thrombin may also be useful for *in vivo* therapeutic purposes, as well as for various *in vitro* applications. In addition, PRO269 polypeptides and portions thereof may have therapeutic use as an antithrombotic agent with reduced risk for hemorrhage as compared with 35 heparin. Peptides having homology to thrombomodulin are particularly desirable.

PRO287 polypeptides and portions thereof which effect the activity of bone morphogenic protein "BMP1"/procollagen C-proteinase (PCP) may also be useful for *in vivo* therapeutic purposes, as well as for various

residues (340-343). Carcinoembryonic antigen precursor, as explained in the Background is a tumor-specific antigen, and as such, is a recognized marker and therapeutic target for the diagnosis and treatment of colon cancer. The expression of tumor-specific antigens is often associated with the progression of neoplastic tissue disorders. Native PRO346 (SEQ ID NO:320) and P_W06874, a human carcinoembryonic antigen CEA-d have a Blast score of 224 and homology of 28% between residues 2 to 343 and 67 to 342, respectively. This homology includes the entire 5 extracellular domain residues of native PRO346, minus the initiator methionine (residues 2 to 18) as well as several transmembrane residues (340-343).

PRO268 polypeptides which have protein disulfide isomerase activity will be useful for many applications where protein disulfide isomerase activity is desirable including, for example, for use in promoting proper disulfide bond formation in recombinantly produced proteins so as to increase the yield of correctly folded protein. Those of 10 ordinary skill in the art will readily know how to employ such PRO268 polypeptides for such purposes.

PRO330 polypeptides of the present invention which possess biological activity related to that of the prolyl 4-hydroxylase alpha subunit protein may be employed both *in vivo* for therapeutic purposes and *in vitro*. Those of ordinary skill in the art will well know how to employ the PRO330 polypeptides of the present invention for such purposes.

15

55. Anti-PRO Polypeptide Antibodies

The present invention further provides anti-PRO polypeptide antibodies. Exemplary antibodies include polyclonal, monoclonal, humanized, bispecific, and heteroconjugate antibodies.

20

A. Polyclonal Antibodies

The anti-PRO polypeptide antibodies may comprise polyclonal antibodies. Methods of preparing polyclonal antibodies are known to the skilled artisan. Polyclonal antibodies can be raised in a mammal, for example, by one or more injections of an immunizing agent and, if desired, an adjuvant. Typically, the immunizing agent and/or adjuvant will be injected in the mammal by multiple subcutaneous or intraperitoneal injections. The immunizing agent 25 may include the PRO polypeptide or a fusion protein thereof. It may be useful to conjugate the immunizing agent to a protein known to be immunogenic in the mammal being immunized. Examples of such immunogenic proteins include but are not limited to keyhole limpet hemocyanin, serum albumin, bovine thyroglobulin, and soybean trypsin inhibitor. Examples of adjuvants which may be employed include Freund's complete adjuvant and MPL-TDM adjuvant (monophosphoryl Lipid A, synthetic trehalose dicorynomycolate). The immunization protocol may be 30 selected by one skilled in the art without undue experimentation.

B. Monoclonal Antibodies

The anti-PRO polypeptide antibodies may, alternatively, be monoclonal antibodies. Monoclonal antibodies may be prepared using hybridoma methods, such as those described by Kohler and Milstein, *Nature*, 256:495 (1975). 35 In a hybridoma method, a mouse, hamster, or other appropriate host animal, is typically immunized with an immunizing agent to elicit lymphocytes that produce or are capable of producing antibodies that will specifically bind to the immunizing agent. Alternatively, the lymphocytes may be immunized *in vitro*.

The immunizing agent will typically include the PRO polypeptide of interest or a fusion protein thereof. Generally, either peripheral blood lymphocytes ("PBLs") are used if cells of human origin are desired, or spleen cells or lymph node cells are used if non-human mammalian sources are desired. The lymphocytes are then fused with an immortalized cell line using a suitable fusing agent, such as polyethylene glycol, to form a hybridoma cell [Goding, Monoclonal Antibodies: Principles and Practice, Academic Press, (1986) pp. 59-103]. Immortalized cell lines are 5 usually transformed mammalian cells, particularly myeloma cells of rodent, bovine and human origin. Usually, rat or mouse myeloma cell lines are employed. The hybridoma cells may be cultured in a suitable culture medium that preferably contains one or more substances that inhibit the growth or survival of the unfused, immortalized cells. For example, if the parental cells lack the enzyme hypoxanthine guanine phosphoribosyl transferase (HGPRT or HPRT), the culture medium for the hybridomas typically will include hypoxanthine, aminopterin, and thymidine 10 ("HAT medium"), which substances prevent the growth of HGPRT-deficient cells.

Preferred immortalized cell lines are those that fuse efficiently, support stable high level expression of antibody by the selected antibody-producing cells, and are sensitive to a medium such as HAT medium. More preferred immortalized cell lines are murine myeloma lines, which can be obtained, for instance, from the Salk Institute Cell Distribution Center, San Diego, California and the American Type Culture Collection, Rockville, 15 Maryland. Human myeloma and mouse-human heteromyeloma cell lines also have been described for the production of human monoclonal antibodies [Kozbor, *J. Immunol.*, 133:3001 (1984); Brodeur *et al.*, Monoclonal Antibody Production Techniques and Applications, Marcel Dekker, Inc., New York, (1987) pp. 51-63].

The culture medium in which the hybridoma cells are cultured can then be assayed for the presence of monoclonal antibodies directed against the PRO polypeptide of interest. Preferably, the binding specificity of 20 monoclonal antibodies produced by the hybridoma cells is determined by immunoprecipitation or by an *in vitro* binding assay, such as radioimmunoassay (RIA) or enzyme-linked immunoabsorbent assay (ELISA). Such techniques and assays are known in the art. The binding affinity of the monoclonal antibody can, for example, be determined by the Scatchard analysis of Munson and Pollard, *Anal. Biochem.*, 107:220 (1980).

After the desired hybridoma cells are identified, the clones may be subcloned by limiting dilution procedures 25 and grown by standard methods [Goding, supra]. Suitable culture media for this purpose include, for example, Dulbecco's Modified Eagle's Medium and RPMI-1640 medium. Alternatively, the hybridoma cells may be grown *in vivo* as ascites in a mammal.

The monoclonal antibodies secreted by the subclones may be isolated or purified from the culture medium or ascites fluid by conventional immunoglobulin purification procedures such as, for example, protein A-Sepharose, 30 hydroxylapatite chromatography, gel electrophoresis, dialysis, or affinity chromatography.

The monoclonal antibodies may also be made by recombinant DNA methods, such as those described in U.S. Patent No. 4,816,567. DNA encoding the monoclonal antibodies of the invention can be readily isolated and sequenced using conventional procedures (e.g., by using oligonucleotide probes that are capable of binding specifically to genes encoding the heavy and light chains of murine antibodies). The hybridoma cells of the invention serve as a preferred source of such DNA. Once isolated, the DNA may be placed into expression vectors, which are then transfected into host cells such as simian COS cells, Chinese hamster ovary (CHO) cells, or myeloma cells that do not otherwise produce immunoglobulin protein, to obtain the synthesis of monoclonal antibodies in the 35

recombinant host cells. The DNA also may be modified, for example, by substituting the coding sequence for human heavy and light chain constant domains in place of the homologous murine sequences [U.S. Patent No. 4,816,567; Morrison et al., *supra*] or by covalently joining to the immunoglobulin coding sequence all or part of the coding sequence for a non-immunoglobulin polypeptide. Such a non-immunoglobulin polypeptide can be substituted for the constant domains of an antibody of the invention, or can be substituted for the variable domains of one antigen-combining site of an antibody of the invention to create a chimeric bivalent antibody.

The antibodies may be monovalent antibodies. Methods for preparing monovalent antibodies are well known in the art. For example, one method involves recombinant expression of immunoglobulin light chain and modified heavy chain. The heavy chain is truncated generally at any point in the Fc region so as to prevent heavy chain crosslinking. Alternatively, the relevant cysteine residues are substituted with another amino acid residue or are deleted so as to prevent crosslinking.

10 *In vitro* methods are also suitable for preparing monovalent antibodies. Digestion of antibodies to produce fragments thereof, particularly, Fab fragments, can be accomplished using routine techniques known in the art.

C. Humanized Antibodies

15 The anti-PRO polypeptide antibodies of the invention may further comprise humanized antibodies or human antibodies. Humanized forms of non-human (e.g., murine) antibodies are chimeric immunoglobulins, immunoglobulin chains or fragments thereof (such as Fv, Fab, Fab', F(ab')₂ or other antigen-binding subsequences of antibodies) which contain minimal sequence derived from non-human immunoglobulin. Humanized antibodies include human immunoglobulins (recipient antibody) in which residues from a complementary determining region (CDR) of the recipient are replaced by residues from a CDR of a non-human species (donor antibody) such as mouse, rat or rabbit having the desired specificity, affinity and capacity. In some instances, Fv framework residues of the human immunoglobulin are replaced by corresponding non-human residues. Humanized antibodies may also comprise residues which are found neither in the recipient antibody nor in the imported CDR or framework sequences. In general, the humanized antibody will comprise substantially all of at least one, and typically two, 20 variable domains, in which all or substantially all of the CDR regions correspond to those of a non-human immunoglobulin and all or substantially all of the FR regions are those of a human immunoglobulin consensus sequence. The humanized antibody optimally also will comprise at least a portion of an immunoglobulin constant region (Fc), typically that of a human immunoglobulin [Jones et al., *Nature*, 321: 522-525 (1986); Riechmann et al., *Nature*, 332:323-329 (1988); and Presta, *Curr. Op. Struct. Biol.*, 2:593-596 (1992)].

25 30 Methods for humanizing non-human antibodies are well known in the art. Generally, a humanized antibody has one or more amino acid residues introduced into it from a source which is non-human. These non-human amino acid residues are often referred to as "import" residues, which are typically taken from an "import" variable domain. Humanization can be essentially performed following the method of Winter and co-workers [Jones et al., *Nature*, 321: 522-525 (1986); Riechmann et al., *Nature*, 332:323-327 (1988); Verhoeyen et al., *Science*, 239:1534-1536 (1988)], 35 by substituting rodent CDRs or CDR sequences for the corresponding sequences of a human antibody. Accordingly, such "humanized" antibodies are chimeric antibodies (U.S. Patent No. 4,816,567), wherein substantially less than an intact human variable domain has been substituted by the corresponding sequence from a non-human species. In

practice, humanized antibodies are typically human antibodies in which some CDR residues and possibly some FR residues are substituted by residues from analogous sites in rodent antibodies.

Human antibodies can also be produced using various techniques known in the art, including phage display libraries [Hoogenboom and Winter, *J. Mol. Biol.*, 227:381 (1991); Marks *et al.*, *J. Mol. Biol.*, 222:581 (1991)]. The techniques of Cole *et al.* and Boerner *et al.* are also available for the preparation of human monoclonal antibodies 5 (Cole *et al.*, *Monoclonal Antibodies and Cancer Therapy*, Alan R. Liss, p. 77 (1985) and Boerner *et al.*, *J. Immunol.*, 147(1):86-95 (1991)).

D. Bispecific Antibodies

Bispecific antibodies are monoclonal, preferably human or humanized, antibodies that have binding 10 specificities for at least two different antigens. In the present case, one of the binding specificities is for the PRO polypeptide, the other one is for any other antigen, and preferably for a cell-surface protein or receptor or receptor subunit.

Methods for making bispecific antibodies are known in the art. Traditionally, the recombinant production 15 of bispecific antibodies is based on the co-expression of two immunoglobulin heavy-chain/light-chain pairs, where the two heavy chains have different specificities [Milstein and Cuello, *Nature*, 305:537-539 (1983)]. Because of the random assortment of immunoglobulin heavy and light chains, these hybridomas (quadromas) produce a potential mixture of ten different antibody molecules, of which only one has the correct bispecific structure. The purification of the correct molecule is usually accomplished by affinity chromatography steps. Similar procedures are disclosed in WO 93/08829, published 13 May 1993, and in Traunecker *et al.*, *EMBO J.*, 10:3655-3659 (1991).

20 Antibody variable domains with the desired binding specificities (antibody-antigen combining sites) can be fused to immunoglobulin constant domain sequences. The fusion preferably is with an immunoglobulin heavy-chain constant domain, comprising at least part of the hinge, CH₂, and CH₃ regions. It is preferred to have the first heavy-chain constant region (CH₁) containing the site necessary for light-chain binding present in at least one of the fusions. DNAs encoding the immunoglobulin heavy-chain fusions and, if desired, the immunoglobulin light chain, are inserted 25 into separate expression vectors, and are co-transfected into a suitable host organism. For further details of generating bispecific antibodies see, for example, Suresh *et al.*, *Methods in Enzymology*, 121:210 (1986).

E. Heteroconjugate Antibodies

Heteroconjugate antibodies are also within the scope of the present invention. Heteroconjugate antibodies 30 are composed of two covalently joined antibodies. Such antibodies have, for example, been proposed to target immune system cells to unwanted cells [U.S. Patent No. 4,676,980], and for treatment of HIV infection [WO 91/00360; WO 92/200373; EP 03089]. It is contemplated that the antibodies may be prepared *in vitro* using known methods in synthetic protein chemistry, including those involving crosslinking agents. For example, immunotoxins 35 may be constructed using a disulfide exchange reaction or by forming a thioether bond. Examples of suitable reagents for this purpose include iminothiolate and methyl-4-mercaptoputyrimidate and those disclosed, for example, in U.S. Patent No. 4,676,980.

56. Uses for Anti-Pro Polypeptide Antibodies

The anti-PRO polypeptide antibodies of the invention have various utilities. For example, anti-PRO polypeptide antibodies may be used in diagnostic assays for a PRO polypeptide, e.g., detecting its expression in specific cells, tissues, or serum. Various diagnostic assay techniques known in the art may be used, such as competitive binding assays, direct or indirect sandwich assays and immunoprecipitation assays conducted in either 5 heterogeneous or homogeneous phases [Zola, Monoclonal Antibodies: A Manual of Techniques, CRC Press, Inc. (1987) pp. 147-158]. The antibodies used in the diagnostic assays can be labeled with a detectable moiety. The detectable moiety should be capable of producing, either directly or indirectly, a detectable signal. For example, the detectable moiety may be a radioisotope, such as ^3H , ^{14}C , ^{32}P , ^{35}S , or ^{125}I , a fluorescent or chemiluminescent compound, such as fluorescein isothiocyanate, rhodamine, or luciferin, or an enzyme, such as alkaline phosphatase, 10 beta-galactosidase or horseradish peroxidase. Any method known in the art for conjugating the antibody to the detectable moiety may be employed, including those methods described by Hunter *et al.*, *Nature*, 144:945 (1962); David *et al.*, *Biochemistry*, 13:1014 (1974); Pain *et al.*, *J. Immunol. Meth.*, 40:219 (1981); and Nygren, *J. Histochem. and Cytochem.*, 30:407 (1982).

Anti-PRO polypeptide antibodies also are useful for the affinity purification of PRO polypeptide from 15 recombinant cell culture or natural sources. In this process, the antibodies against the PRO polypeptide are immobilized on a suitable support, such as a Sephadex resin or filter paper, using methods well known in the art. The immobilized antibody then is contacted with a sample containing the PRO polypeptide to be purified, and thereafter the support is washed with a suitable solvent that will remove substantially all the material in the sample except the PRO polypeptide, which is bound to the immobilized antibody. Finally, the support is washed with another suitable 20 solvent that will release the PRO polypeptide from the antibody.

With regard to PRO211 and PRO217, therapeutic indications include disorders associated with the preservation and maintenance of gastrointestinal mucosa and the repair of acute and chronic mucosal lesions (e.g., enterocolitis, Zollinger-Ellison syndrome, gastrointestinal ulceration and congenital microvillus atrophy), skin diseases associated with abnormal keratinocyte differentiation (e.g., psoriasis, epithelial cancers such as lung squamous cell 25 carcinoma, epidermoid carcinoma of the vulva and gliomas).

With regard to anti-PRO187 antibodies, FGF-8 has been implicated in cellular differentiation and embryogenesis, including the patterning which appears during limb formation. FGF-8 and the PRO187 molecules of the invention therefore are likely to have potent effects on cell growth and development. Diseases which relate to cellular growth and differentiation are therefore suitable targets for therapeutics based on functionality similar to 30 FGF-8. For example, diseases related to growth or survival of nerve cells including Parkinson's disease, Alzheimer's disease, ALS, neuropathies. Additionally, disease related to uncontrolled cell growth, e.g., cancer, would also be expected therapeutic targets.

Native PRO533 is a 216 amino acid polypeptide of which residues 1-22 are the signal sequence. Residues 3 to 216 have a Blast score of 509, corresponding to 53% homology to fibroblast growth factor. At the nucleotide 35 level, DNA47412, the EST from which PCR oligos were generated to isolate the full length DNA49435-1219, has been observed to map to 11p15. Sequence homology to the 11p15 locus would indicate that PRO533 may have utility in the treatment of Usher Syndrome or Atrophy areata.

2) for use as probes to isolate a clone of the full-length coding sequence for PRO266.

A pair of PCR primers (forward and reverse) were synthesized:

forward PCR primer 5'-GTTGGATCTGGCAACAATAAC-3' (SEQ ID NO:92)

reverse PCR primer 5'-ATTGTTGTGCAGGCTGAGTTAAG-3' (SEQ ID NO:93)

Additionally, a synthetic oligonucleotide hybridization probe was constructed which had the following nucleotide

5 sequence

hybridization probe

5'-GGTGGCTATACATGGATAGCAATTACCTGGACACGCTGTCCCGGG-3' (SEQ ID NO:94)

In order to screen several libraries for a source of a full-length clone, DNA from the libraries was screened by PCR amplification with the PCR primer pair identified above. A positive library was then used to isolate clones

10 encoding the PRO266 gene using the probe oligonucleotide and one of the PCR primers.

RNA for construction of the cDNA libraries was isolated from human fetal brain tissue. DNA sequencing of the clones isolated as described above gave the full-length DNA sequence for PRO266 [herein designated as UNQ233 (DNA37150-1178)] (SEQ ID NO:90) and the derived protein sequence for PRO266.

The entire nucleotide sequence of UNQ233 (DNA37150-1178) is shown in Figure 33 (SEQ ID NO:90).

15 Clone UNQ233 (DNA37150-1178) contains a single open reading frame with an apparent translational initiation site at nucleotide positions 167-169 and ending at the stop codon after nucleotide position 2254 of SEQ ID NO:90. The predicted polypeptide precursor is 696 amino acids long (Figure 34). Clone UNQ233 (DNA37150-1178) has been deposited with ATCC and is assigned ATCC deposit no. ATCC 209401.

Analysis of the amino acid sequence of the full-length PRO266 polypeptide suggests that portions of it 20 possess significant homology to the SLIT protein, thereby indicating that PRO266 may be a novel leucine rich repeat protein.

EXAMPLE 15: Isolation of cDNA Clones Encoding Human PRO269

A consensus DNA sequence was assembled relative to other EST sequences using phrap as described in 25 Example 1 above. This consensus sequence is herein designated DNA35705. Based on the DNA35705 consensus sequence, oligonucleotides were synthesized: 1) to identify by PCR a cDNA library that contained the sequence of interest, and 2) for use as probes to isolate a clone of the full-length coding sequence for PRO269.

Forward and reverse PCR primers were synthesized:

forward PCR primer (.f1) 5'-TGGAAGGAGATGCGATGCCACCTG -3'

30 (SEQ ID NO:97)

forward PCR primer (.f2) 5'-TGACCAGTGGGAAGGACAG-3' (SEQ ID NO:98)

forward PCR primer (.f3) 5'-ACAGAGCAGAGGGTGCCTTG-3' (SEQ ID NO:99)

reverse PCR primer (.r1) 5'-TCAGGGACAAGTGGTGTCTCTCCC-3'

(SEQ ID NO:100)

35 reverse PCR primer (.r2) 5'-TCAGGGAAGGAGTGTGCAGTTCTG-3'

(SEQ ID NO:101)

Additionally, a synthetic oligonucleotide hybridization probe was constructed from the consensus DNA35705 sequence which had the following nucleotide sequence:

hybridization probe

5'-ACAGCTCCGATCTCAGTTACTGCATCGCGACGAAATGGCGCTCGCT-3' (SEQ ID NO:102)

- In order to screen several libraries for a source of a full-length clone, DNA from the libraries was screened
5 by PCR amplification with the PCR primer pairs identified above. A positive library was then used to isolate clones encoding the PRO269 gene using the probe oligonucleotide and one of the PCR primers.

RNA for construction of the cDNA libraries was isolated from human fetal kidney tissue.

DNA sequencing of the clones isolated as described above gave the full-length DNA sequence for PRO269 [herein designated as UNQ236 (DNA38260-1180)] (SEQ ID NO:95) and the derived protein sequence for PRO269.

- 10 The entire nucleotide sequence of UNQ236 (DNA38260-1180) is shown in Figure 35 (SEQ ID NO:95). Clone UNQ236 (DNA38260-1180) contains a single open reading frame with an apparent translational initiation site at nucleotide positions 314-316 and ending at the stop codon at nucleotide positions 1784-1786 (Fig. 35; SEQ ID NO:95). The predicted polypeptide precursor is 490 amino acids long (Fig. 36). Clone UNQ236 (DNA38260-1180) has been deposited with ATCC and is assigned ATCC deposit no. ATCC 209397.

- 15 Analysis of the amino acid sequence of the full-length PRO269 suggests that portions of it possess significant homology to the human thrombomodulin proteins, thereby indicating that PRO269 may possess one or more thrombomodulin-like domains.

EXAMPLE 16: Isolation of cDNA Clones Encoding Human PRO287

- 20 A consensus DNA sequence encoding PRO287 was assembled relative to the other identified EST sequences as described in Example 1 above, wherein the consensus sequence is designated herein as DNA28728. Based on the DNA28728 consensus sequence, oligonucleotides were synthesized to identify by PCR a cDNA library that contained the sequence of interest and for use as probes to isolate a clone of the full-length coding sequence for PRO287.

A pair of PCR primers (forward and reverse) were synthesized:

- 25 forward PCR primer 5'-CCGATTCTCATAGACCTCGAGAGT-3' (SEQ ID NO:105)

- reverse PCR primer 5'-GTCAAGGAGTCCTCCACAATAC-3' (SEQ ID NO:106)

Additionally, a synthetic oligonucleotide hybridization probe was constructed from the consensus DNA28728 sequence which had the following nucleotide sequence

hybridization probe

- 30 5'-GTGTACAATGGCCATGCCAATGGCCAGCGCATTGGCCGTTCTGT-3'

(SEQ ID NO:107)

In order to screen several libraries for a source of a full-length clone, DNA from the libraries was screened by PCR amplification with the PCR primer pair identified above. A positive library was then used to isolate clones encoding the PRO287 gene using the probe oligonucleotide and one of the PCR primers.

- 35 RNA for construction of the cDNA libraries was isolated from human fetal kidney tissue.

DNA sequencing of the clones isolated as described above gave the full-length DNA sequence for PRO287 [herein designated as UNQ250 (DNA39969-1185), SEQ ID NO:103] and the derived protein sequence for PRO287.

in epithelial cells of the fetal adrenal cortex. All adult tissues were negative.

Fetal tissues examined (E12-E16 weeks) include: Placenta, umbilical cord, liver, kidney, adrenals, thyroid, lungs, heart, great vessels, oesophagus, stomach, small intestine, spleen, thymus, pancreas, brain, eye, spinal cord, body wall, pelvis and lower limb.

- 5 Adult tissues examined: Liver, kidney, adrenal, spleen, lymph node, pancreas, lung, skin, cerebral cortex (rm), hippocampus(rm), cerebellum(rm), bladder, stomach, colon and colonic carcinoma. Acetominophen induced liver injury and hepatic cirrhosis.

A secondary screen evidenced expression over stromal mononuclear cells probably histiocytes.

(14) DNA38268-1188 (PRO295)

- 10 High expression over ganglion cells in human fetal spinal ganglia and over large neurones in the anterior horns of the developing spinal cord. In the adult there is expression in the chimp adrenal medulla (neural), neurones of the rhesus monkey brain (hippocampus [+ + +] and cerebral cortex) and neurones in ganglia in the normal adult human prostate (the only section that contains ganglion cells, ie expression in this cell type is presumed NOT to be confined to the prostate). All other tissues negative.
- 15 Human fetal tissues examined (E12-E16 weeks) include: Placenta, umbilical cord, liver, kidney, adrenals, thyroid, lungs, great vessels, stomach, small intestine, spleen, thymus, pancreas, brain, eye, spinal cord, body wall, pelvis, testis and lower limb.

Adult human tissues examined: Kidney (normal and end-stage), adrenal, spleen, lymph node, pancreas, lung, eye (inc. retina), bladder, liver (normal, cirrhotic, acute failure).

- 20 Non-human primate tissues examined:

Chimp Tissues: adrenal

Rhesus Monkey Tissues: Cerebral cortex, hippocampus, cerebellum.

Deposit of Material

- 25 The following materials have been deposited with the American Type Culture Collection, 12301 Parklawn Drive, Rockville, MD, USA (ATCC):

	<u>Material</u>	<u>ATCC Dep. No.</u>	<u>Deposit Date</u>
	DNA32292-1131	ATCC 209258	September 16, 1997
	DNA33094-1131	ATCC 209256	September 16, 1997
30	DNA33223-1136	ATCC 209264	September 16, 1997
	DNA34435-1140	ATCC 209250	September 16, 1997
	DNA27864-1155	ATCC 209375	October 16, 1997
	DNA36350-1158	ATCC 209378	October 16, 1997
	DNA32290-1164	ATCC 209384	October 16, 1997
35	DNA35639-1172	ATCC 209396	October 17, 1997
	DNA33092-1202	ATCC 209420	October 28, 1997
	DNA49435-1219	ATCC 209480	November 21, 1997
	DNA35638-1141	ATCC 209265	September 16, 1997
	DNA32298-1132	ATCC 209257	September 16, 1997
40	DNA33089-1132	ATCC 209262	September 16, 1997
	DNA33786-1132	ATCC 209253	September 16, 1997
	DNA35918-1174	ATCC 209402	October 17, 1997

	DNA37150-1178	ATCC 209401	October 17, 1997
	DNA38260-1180	ATCC 209397	October 17, 1997
	DNA39969-1185	ATCC 209400	October 17, 1997
	DNA32286-1191	ATCC 209385	October 16, 1997
	DNA33461-1199	ATCC 209367	October 15, 1997
5	DNA40628-1216	ATCC 209432	November 7, 1997
	DNA33221-1133	ATCC 209263	September 16, 1997
	DNA33107-1135	ATCC 209251	September 16, 1997
	DNA35557-1137	ATCC 209255	September 16, 1997
	DNA34434-1139	ATCC 209252	September 16, 1997
10	DNA33100-1159	ATCC 209373	October 16, 1997
	DNA35600-1162	ATCC 209370	October 16, 1997
	DNA34436-1238	ATCC 209523	December 10, 1997
	DNA33206-1165	ATCC 209372	October 16, 1997
	DNA35558-1167	ATCC 209374	October 16, 1997
15	DNA35599-1168	ATCC 209373	October 16, 1997
	DNA36992-1168	ATCC 209382	October 16, 1997
	DNA34407-1169	ATCC 209383	October 16, 1997
	DNA35841-1173	ATCC 209403	October 17, 1997
	DNA33470-1175	ATCC 209398	October 17, 1997
20	DNA34431-1177	ATCC 209399	October 17, 1997
	DNA39510-1181	ATCC 209392	October 17, 1997
	DNA39423-1182	ATCC 209387	October 17, 1997
	DNA40620-1183	ATCC 209388	October 17, 1997
	DNA40604-1187	ATCC 209394	October 17, 1997
25	DNA38268-1188	ATCC 209421	October 28, 1997
	DNA37151-1193	ATCC 209393	October 17, 1997
	DNA35673-1201	ATCC 209418	October 28, 1997
	DNA40370-1217	ATCC 209485	November 21, 1997
	DNA42551-1217	ATCC 209483	November 21, 1997
30	DNA39520-1217	ATCC 209482	November 21, 1997
	DNA41225-1217	ATCC 209491	November 21, 1997
	DNA43318-1217	ATCC 209481	November 21, 1997
	DNA40587-1231	ATCC 209438	November 7, 1997
	DNA41338-1234	ATCC 209927	June 2, 1998
35	DNA40981-1234	ATCC 209439	November 7, 1997
	DNA37140-1234	ATCC 209489	November 21, 1997
	DNA40982-1235	ATCC 209433	November 7, 1997
	DNA41379-1236	ATCC 209488	November 21, 1997
	DNA44167-1243	ATCC 209434	November 7, 1997
40	DNA39427-1179	ATCC 209395	October 17, 1997
	DNA40603-1232	ATCC 209486	November 21, 1997
	DNA43466-1225	ATCC 209490	November 21, 1997
	DNA43046-1225	ATCC 209484	November 21, 1997
	DNA35668-1171	ATCC 209371	October 16, 1997

45

These deposit were made under the provisions of the Budapest Treaty on the International Recognition of the Deposit of Microorganisms for the Purpose of Patent Procedure and the Regulations thereunder (Budapest Treaty). This assures maintenance of a viable culture of the deposit for 30 years from the date of deposit. The deposits will be made available by ATCC under the terms of the Budapest Treaty, and subject to an agreement between Genentech, Inc. and ATCC, which assures permanent and unrestricted availability of the progeny of the culture of the deposit to the public upon issuance of the pertinent U.S. patent or upon laying open to the public of any U.S. or foreign patent application, whichever comes first, and assures availability of the progeny to one determined

by the U.S. Commissioner of Patents and Trademarks to be entitled thereto according to 35 USC § 122 and the Commissioner's rules pursuant thereto (including 37 CFR § 1.14 with particular reference to 886 OG 638).

The assignee of the present application has agreed that if a culture of the materials on deposit should die or be lost or destroyed when cultivated under suitable conditions, the materials will be promptly replaced on notification with another of the same. Availability of the deposited material is not to be construed as a license to practice the invention in contravention of the rights granted under the authority of any government in accordance with its patent laws.

The foregoing written specification is considered to be sufficient to enable one skilled in the art to practice the invention. The present invention is not to be limited in scope by the construct deposited, since the deposited embodiment is intended as a single illustration of certain aspects of the invention and any constructs that are functionally equivalent are within the scope of this invention. The deposit of material herein does not constitute an admission that the written description herein contained is inadequate to enable the practice of any aspect of the invention, including the best mode thereof, nor is it to be construed as limiting the scope of the claims to the specific illustrations that it represents. Indeed, various modifications of the invention in addition to those shown and described herein will become apparent to those skilled in the art from the foregoing description and fall within the scope of the appended claims.

FIGURE 35

AGTCGACTGCGTCCCTGTACCCGGCGCCAGCTGTGTTCTGACCCCAGAATAACTCAGG
 GCTGCACCAGGGCTGGCAGCGCTCCGCACACATTCTCTGTCGCGGCCAAGGGAAACTGT
 TGGCCGCTGGGCCCGCGGGGGATTCTTGGCAGTTGGGGGTCCTGCGGGAGCGAGGGCG
 GAGGGGAAGGGAGGGGAACCGGGTTGGGAAGCCAGCTGTAGAGGGCGGTGACCGCGCT
 CCAGACACAGCTCTGCGTCCTCGAGCGGGACAGATCCAAGTTGGAGCAGCTCTGCGTGC
 GGGCCTCAGAGA
 ><MET {trans=1-s, dir=f, res=1}
 ATGAGGCCGGCGTTCGCCCTGTGCCTCTGGCAGGCCTCTGGCCCGGGCCGGCGGC
 GGCAGAACACCCCACTGCCGACCGTGCCTGGCTCGGCCCTCGGGGCCTGCTACAGCCTG
 CACCACGCTACCATGAAGCGGCAGGCCGAGGAGGCCTGCATCCTGCGAGGTGGGCG
 CTCAGCACCGTGCCTGCCGGCGAGCTGCCTGCTCGCTCCCTGCGGGCAGGC
 CCAGGGCCGGAGGGGCTCAAAGACCTGCTGTTCTGGCTGCAGTGAGCGCAGGC
 TCCCACGTGACCCCTGGAGAACGAGCTTGCCTGGGGTTCTCTGGCTGTCCTCCGACCC
 GGCCTCGAAAGCGACACGCTGCAGTGGTGGAGGAGCCCCAACGCTCCTGCAACCGCG
 CGGAGATGCGCGGTACTCCAGGCCACCGTGGCTCGAGCCGCAGGCTGGAAGGAGATG
 CGATGCCACCTGCGCGCCAACGGCTACCTGTGCAAGTACCAAGTTGAGGTCTGTGTCCT
 GCGCCGCGCCCCGGGCGCCTAACTTGAGCTATCGCGCCCTTCCAGCTGCACAGC
 GCCGCTCTGGACTTCAGTCACCTGGACCGAGGTGAGTGCCTCTGCCGGGACAGCTC
 CCGATCTCAGTTACTTGATCGCGGACGAAATGGCGCTCGCTGGACAAACTCTGGG
 GATGTGTTGTCCTGCCCTGGGGAGGTACCTCGTGCTGGCAAATGCGCAGAGCTCC
 AACTGCCTAGACGACTTGGGAGGCTTGCGAATGTGCTACGGCTCGAGCTGGG
 AAGGACGGCCGCTTGTGACCAGTGGGAAGGACAGCCGACCCCTGGGGGACCGGG
 GTGCCCAACAGCGCCGCCACTGCAACCAGCCCGTGGCGAGAGAACATGCCA
 ATCAGGGTGCACGAGAACGACTGGGAGAGACACCACCTGTCCTGAACAAGACAATT
 ACAGTAACATCTATTCTGAGATTCTCGATGGGATCACAGAGCAGATGTCTACCC
 TCCCTCAAGCCGAGTCAAAGGCCACTATCACCCATCAGGGAGCGTATTCCAAG
 ATTCTACGACTCCTCTGCCACTCCTCAGGCTTGACTCCTCTGCCGTGGCTTC
 ATATTGTGAGCACAGCAGTAGTAGTGTTGGTGATCTGACAGTACTGGGCTT
 GTCAAGCTTGCTTACGAAAGCCCTTCCAGCCAAGGAAGGAGTCTATGGGCC
 CCGGGCTGGAGAGTGATCCTGAGCCGCTGCTTGGGCTCCAGTCTGCACATT
 GCACAACAAATGGGGTGAAGTCGGGACTGTGATCTGCGGGACAGAGCAGAGGG
 GCCTGCTGCGAGTCCCTTGGCTCTAGTGATGCATAGGAAACAGGGACATGG
 GAAACAGTTTCACTTTGATGAAACGGGAACCAAGAGGAACCTACTTGTA
 ACTGACATTCTGCAGAAATCCCCCTTCTCTAAATTCCCTTACTCC
 ACTGAGGAGCTAAATCA
 GAACCTGCACACTCCTCCCTGATGATAGAGGAAGTGGAAAGTGCCTT
 TAGGATGGTGATAC
 TGGGGACGGGTAGTGCTGGGGAGAGATATTCTTATGTTATT
 CGGAGAATTGGAG
 AAGTGATTGAACTTTCAAGACATTGAAACAAATAGAACACA
 AAATATAATTACATTAAA
 AAAATAATTCTACCAAAATGGAAAGGAAATGTTCTATGTT
 CAGGCTAGGAGTATATT
 GGTCGAAATCCCAGGGAAAAAATAAAAATTAAAGGATTGAT

FIGURE 36

MRPAFALCLLWQALWPAGPGGGEHPTADRAGCSASGACYSLHHATMKRQAAEEACILRGGA
LSTVRAGAELRAVLALLRAGPGPGGGSKDLLFWVALERRSHCTLENEPLRGFSWLSSDP
GGLESDTLQWVEEPQRSCARRCAVLQATGGVEPAGWKEMRCHLRANGYLCKYQFEVLCP
APRPGAASNLSYRAPFQLHSAALDFSPPGTEVSALCRGQLPISVTCIADEIGARWDKLSG
DVLCPCPGRYLРАГКАЕLPNCLDDLGGFACECATGFELGKDGRSCVTSGEGOPTLGGTG
VPTRRPPATATSPVPQRTWPIRVDEKLGETPLVPEQDNSVTSIPEIPRWGSQSTMSTLQM
SLQAESKATITPSGSVISKFNSTTSSATPQAFDSSSAVVFIFVSTAVVVLVILTMVLGL
VKLCFHESPSSQPRKESMGPPGLESDPPEPAALGSSAHCTNNGVKGDCDLRDRAEGALL
AESPLGSSDA